

## Development and Characterization of a Paper-Based Sensor for Colorimetric Urea Detection in Artificial Urine

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**Abstract** – Toxic nitrogen, which is formed as a result of the breakdown of proteins, is converted into non-toxic urea by the enzymes in the "urea cycle". While a small part of the produced urea is eliminated with sweat, a large part is removed from the body by the kidneys with urine. Therefore, urea determination in blood and urine provides important information about kidney functions. In this study, a colorimetric paper-based sensor was developed and characterized for use in urea detection. Basically, urea measurement was carried out by immobilization of urease enzyme on pH sensitive papers. The determination of urea was made by observing the changes in pH caused by ammonia with basic properties, which is formed as a result of the destruction of urea by urease. In the presence of different concentrations of urea, the colorimetric changes on the sensor were visualized with a camera and analyzed with the ImageJ program in the computer environment. In the first stage, a calibration curve showing the relationship between urea concentration and color intensity was obtained, and then the detection limit and sensitivity of the system were determined from this curve. Finally, the selectivity of the sensor was demonstrated by testing the different molecules involved.

**Keywords** – Paper-Based Sensor, Urea, Urine, Colorimetric Sensor

### I. INTRODUCTION

Ammonia (NH<sub>3</sub>) is formed as a result of deamidation and deamination reactions in mammals. NH<sub>3</sub> is a very strong water-soluble base and can raise the pH of the living environment to a harmful level. The ammonia formed, together with the CO<sub>2</sub> present in the blood as a product of metabolism, is converted by the liver into a non-toxic compound, urea. 90% of the urea formed is excreted by the kidneys in the urine, and the rest is excreted through the digestive system and skin. In a normal adult human, approximately 30 grams of urea is excreted in 24 hours. The urea excreted in the urine depends on the protein ratio, the urea synthesis rate in the liver, and the excretion rate of urea from the kidneys. Determination of urea has an important place for biomedical and clinical applications. An increase in urea concentration in the blood or a decrease in the urine is an important indicator of

kidney dysfunction. (Hamlaoui et al. 2002). In this study, a paper-based biosensor capable of sensitive and selective urea detection was developed and characterized. The determination of urea was made by observing the changes in pH caused by basic ammonia formed as a result of the degradation of urea by urease. Litmus paper, which can respond to basic pH, was used in the construction of the sensor. Paper-based colorimetric sensors were produced by immobilizing the urease enzyme on this paper by physical adsorption technique. The sensors were tested with artificial urine samples containing urea at different concentrations, and the colorimetric changes that occurred as a result of the detection were visualized with a camera and analyzed with the ImageJ program in the computer environment. The results obtained revealed the potential of the developed sensor to be used in real samples.

## II. MATERIALS AND METHOD

### A. Design and Fabrication of $\mu$ PADs

The Microsoft PowerPoint application was used to create the design for  $\mu$ PAD. At the bottom of the detection area, a thin hydrophilic channel was created, and the edges of the detection area were constructed with larger and longer hydrophobic barriers. The intended  $\mu$ PADs were printed using the wax printer on Whatman filter paper. After printing,  $\mu$ PADs were placed on a heater at  $+120^{\circ}\text{C}$  for roughly 3–4 minutes to allow the solid ink to penetrate the chromatography paper's pores and thus form the hydrophobic barriers. To ensure a uniform heat transfer, the heater was covered with aluminium foil, and the paper was weighed down by a few kilograms. Next, 2.5  $\mu\text{L}$  of urease solution was added to the detection area and allowed to dry for 10 minutes at room temperature. A litmus paper was placed on top of each  $\mu$ PAD and the two was sandwiched between two transparent tapes.

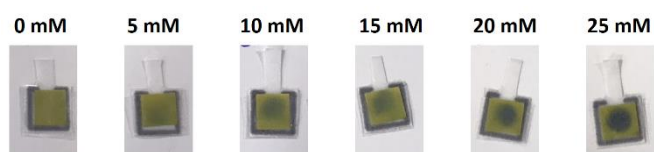


Fig. 1. Color change observed in the detection area of  $\mu$ PADs with artificial urine solutions containing urea at different concentrations.

### B. Colorimetric Detection of Urea in Artificial Urine and Selectivity Test

Artificial urine solutions (120.9 mg NaCl, 16.5 mg  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ , 9 mg  $\text{CaCl}_2$  in 15 mL  $\text{dH}_2\text{O}$ , pH: 6.3) containing varying concentrations of urea (0, 5, 10,

15, 20 and 25 mM) were prepared. The sampling pad of  $\mu$ PADs were immersed into these urine solutions and the color change was recorded with a smartphone camera at different time points (0, 2, 4, 6, 8, 10 and 12 min). The images were then analyzed using ImageJ on a computer. A selectivity test was also carried out using various interferents (NaCl, glucose, sucrose, uric acid, dopamine, lactic acid, KCl, and urea) at 1 mM.

## III. RESULTS & DISCUSSION

The fabrication procedure allowed us to make  $\mu$ PADs successfully. The design resulted in no liquid leakage and successful transfer of sample to detection areas for color change. Considering the amount of chemicals used for testing, the cost of analysis was considerably low. Once the  $\mu$ PADs were dipped into artificial urine sample with urea at different concentration, color change was observed in the detection areas of  $\mu$ PADs (Fig. 1). Basically, the enzyme urease catalyzes the conversion of urea to  $\text{CO}_2$  and  $\text{NH}_3$  which is a basic molecule that promptly changes the color of the litmus paper. The color change was imaged at different time intervals and analyzed using ImageJ software. The RGB intensity (weighted) of the images were used for analysis. As can be seen in Fig 2a, the highest RGB intensity was observed at 2 min. Next, the RGB intensity data of all samples with different concentrations of urea were used to obtain a calibration curve. As seen in Fig 2b, the relationship between the urea concentration and RGB intensity data was linear in the range of 5 to 25 mM with an  $R^2$  value of 0.992. The limit of detection value of the sensor was calculated to be 2.39 mM ( $\text{LOD} = 3.3 \sigma/\text{slope}$ ). In addition, the selectivity test proved the

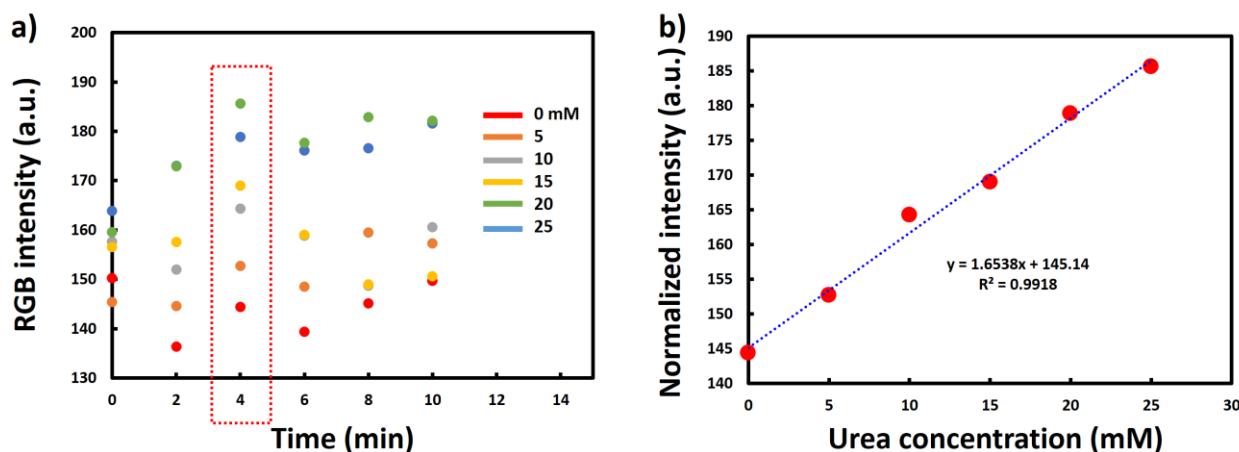


Fig. 2. Color change intensity at different time points (a) along with a calibration curve (b).

specificity of the  $\mu$ PADs for urea detection. Only when urea was present, the  $\mu$ PADs showed a noticeable color change, demonstrating their great potential for selective detection of urea.

#### IV. CONCLUSION

In conclusion, colorimetric detection of urea with  $\mu$ PADs in artificial urine showed promising results. The sensor had a low LOD and a linear response in a relevant concentration range. The selectivity test confirmed the urea detection specificity of the  $\mu$ PADs. The sensor thus has a great potential to be applied to real samples for diagnostic purposes.

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